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## **CHARACTERISTICS OF MIRNA INTERACTION WITH 5'UTR, CDS, 3'UTR mRNA CANDIDATE GENES OF METABOLIC SYNDROME DISEASE**

Studying of the pathogenesis of metabolic syndrome is one of the most topical because of the widespread of this pathology. The study of genetic nature of cardiovascular diseases is one of the most promising areas of molecular medicine worldwide. Identification of a group of individual genetic markers allows early diagnosis and optimization of primary and secondary prevention of metabolic syndrome. Therefore, the process of biomarkers discovery should improve the therapy and assessment of the risk of cardiovascular disease. The search for effective markers is the subject of ongoing research. The miRNAs interaction with mRNAs of candidate genes were predicted using the MirTarget program. The genes responsible for the development of metabolic syndrome, regulated by miRNAs were selected by searching in the PubMed database. ADRA2A and SCAP genes are regulated by 12 and 15 miRNAs through 5'UTR with the highest binding energy is -144 kJ/mole and -151 kJ/mole, respectively. AR, CEBPA, IGFBP2, KL and SIRT1 genes are regulated through CDS by 7, 19, 11, 13 and 6 miRNAs, with the highest binding energy is -134 kJ / mole, -142 kJ/mole, -140 kJ/mole, -142 kJ/mole and -138 kJ/mole, respectively. Clusters of miRNA binding sites with overlapping nucleotide sequences were detected in mRNA of several genes. Several associations of miRNA and genes are proposed as biomarkers for developing methods for diagnosing the metabolic syndrome.

**Key words:** miRNA, mRNA, metabolic syndrome, candidate genes.

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### **miRNA-дың метаболитикалық синдром ауруы кандидатты гендерінің mRNA-дың 5'UTR, CDS және 3'UTR-мен өзара әрекеттің сипаттамалары**

Метаболитикалық синдромының кең таралғандағынан оның патогенезін зерттеуі ең маңыздысы болып табылады. Әлемдік молекулярлы медицинада перспективті түрде және жан-жақты қарастырылып жатқан аурулардың бірі – жүрек-қан тамыр ауруының генетикасын зерттеу. Бөлек генетикалық маркерлерді анықтау – метаболитикалық синдромың біріншілік және екіншілік профилактикасының оптимизациясы мен ерте диагностикасына мүмкіндік береді. Осылайша, биомаркерлер арқылы жүрек-қан тамыр ауруларының қауіп-қатерлерін бағалау және терапиясын жақсартуға болады. Жұмыстың мақсаты – эффективті маркерлерді табу болып табылады. miRNA-лар және кандидатты гендер mRNA-дың арасындағы өзара әрекеттесулері MirTarget бағдарлама арқылы табылады. miRNA-мен реттелінетін метаболитикалық синдромға жауапты гендер PubMed деректер қорынан іздестірілді. ADRA2A және SCAP гендері 5'UTR-де 12 және 15 miRNAs-мен байланысады, жоғары байланысу энергиясы 144 кДж/моль және 151 кДж/моль құрайды. AR, CEBPA, IGFBP2, KL және SIRT1 гендері CDS-те, 7, 19, 11, 13 және 6 miRNAs-мен байланысады, жоғары байланысу энергиясы -134 кДж/моль, -142 кДж/моль, -140 кДж/моль, -142 кДж/моль және 138 кДж/мольді құрайды. Бұл гендерде miRNA-мен байланысатын көптеген

сайттары бар және де осы байланысу кластерлер метаболитикалық синдром диагностикасында биомаркерлер ретінде бола алады.

**Түйін сөздер:** miRNA, mRNA, метаболитикалық синдром, кандидатты гендер.

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### Характеристики взаимодействия miRNA с 5'UTR, CDS и 3'UTR mRNA кандидатных генов метаболического синдрома

Изучение генетической природы метаболического синдрома является одной из наиболее перспективных областей молекулярной медицины во всем мире. Выявление генетических маркеров позволяет разработать раннюю диагностику заболевания и оптимизировать профилактику метаболического синдрома. Обнаружение биомаркеров должно улучшить терапию и оценку риска сердечно-сосудистых заболеваний. Поиск эффективных маркеров метаболического синдрома является предметом текущих исследований. Взаимодействие miRNA с mRNA кандидатных генов было предсказано с помощью программы MirTarget. Гены, ответственные за развитие метаболического синдрома, регулируемые микроРНК, были отобраны путем поиска в базе PubMed. Гены ADRA2A и SCAP регулируются 12 и 15 miRNAs в 5'UTR, наибольшая энергия связывания составляет -144 кДж/моль и -151 кДж/моль соответственно. В CDS mRNA генов AR, CEBPA, IGFBP2, KL и SIRT1 связываются с 7, 19, 11, 13 и 6 miRNA со свободной энергией взаимодействия равной -134 кДж/моль, -142 кДж/моль, -140 кДж/моль, -142 кДж/моль и -138 кДж/моль, соответственно. В mRNA нескольких генов выявлены кластеры сайтов связывания miRNA с наложением нуклеотидных последовательностей. Несколько ассоциаций miRNA и генов предлагаются в качестве биомаркеров для разработки методов диагностики метаболического синдрома.

**Ключевые слова:** miRNA, mRNA, метаболический синдром, гены кандидаты.

## Introduction

Cardiovascular diseases are the leading cause of death worldwide. Essential hypertension is a major risk factor for the development of other cardiovascular diseases and is caused by a combination of environmental and genetic factors, with up to 50% of blood pressure variance currently attributed to an individual's genetic makeup. [1]. Metabolic syndrome is a cluster of various combinations of metabolic abnormalities associated with an increased risk of cardiovascular disease and type 2 diabetes mellitus (DM) [2, 3]. Efforts to date have identified several candidate genes involved in primary hypertension, including *ADD1*, *ABCB1*, *CYP3A5*, *AGT*, *GRK4*, *GNB3* and *NOS3* [4-7]. Genes *ACACB*, *APOA1*, *APOA2*, *APOB*, *APOC1*, *APOC3*, *APOD* and *APOE* are associated with metabolic syndrome [8]. MiRNAs are small noncoding RNAs that have emerged as important regulators of many biological and pathological processes, including those relevant to the development of the heart and cardiovascular disease, such as metabolic syndrome. There are still several interesting reviews about recent progress on miRNAs and cardiovascular disease, which discusses the relationship between miRNAs and cardiac development, myocardial

regeneration, and cardiovascular disease [9-21]. The search for effective markers of the risk of metabolic syndrome is the subject of this research.

## Materials and methods

The nucleotide sequences of candidate genes of the metabolic syndrome (MS) were downloaded from GenBank [<http://www.ncbi.nlm.nih.gov>]. 3701 miRNAs were taken from the publication of Londin E. et al. [22]. The miRNAs binding sites in 5'-untranslated regions (5'UTRs), coding domain sequences (CDSs) and 3'-untranslated regions (3'UTRs) of several genes were predicted using the MirTarget program [23]. This program defines the following features of binding: a) the origin of the initiation of miRNA binding to mRNAs; b) the localization of miRNA binding sites in the 5'UTRs, CDSs and the 3'UTRs of the mRNAs; c) the free energy of hybridization ( $\Delta G$ , kJ/mole); and d) the schemes of nucleotide interactions between miRNAs and mRNAs. The ratio  $\Delta G/\Delta G_m$  (%) was determined for each site ( $\Delta G_m$  equals the free energy of miRNA binding with its perfect complementary nucleotide sequence). The miRNA binding sites located on the mRNAs had  $\Delta G/\Delta G_m$  ratios of 90% or more. The program identifies the positions of the

binding sites on the mRNA, beginning from the first nucleotide of the mRNA's 5'UTR. The MirTarget program found hydrogen bonds between adenine (A) and uracil (U), guanine (G) and cytosine (C), G and U, and A and C. The distances between A and C were equal to those between G and C, A and U, and G and U [24-25]. The numbers of hydrogen bonds in the G-C, A-U, G-U and A-C interactions were found to be 3, 2, 1 and 1, respectively.

## Results and Discussion

miRNAs bound with the high  $\Delta G/\Delta G_m$  value in 5'UTR, CDS and 3'UTR of mRNAs genes participating in MS. 41 target genes were associated with 70 miRNAs in 5'UTR (Table 1).

mRNA of *ADRA2A* gene had binding sites for miR-19-33623-3p, miR-1-2121-3p, miR-19-21199-

3p, miR-17-41168-3p, miR-4-11421-3p, miR-22-16963-5p, miR-8-24509-3p and miR-20-22562-3p located in a cluster from 256 nt to 288 nt. The total length of the miRNAs is 180 nt, which would sequentially take 965 nt of the 5'UTR of this gene. Thus, such arrangement of nine sites in a cluster of 32 nt in length makes sense to reduce the proportion of binding sites.

30 binding sites for miR-19-37933-5p, miR-1-2121-3p, miR-3-8100-5p, miR-19-21199-3p, miR-19-33623-3p, miR-1-155-3p, miR-1-1714-3p, miR-17-40348-5p, miR-2-3313-3p, miR-15-32047-5p, miR-9-28523-5p, miR-2-4453-3p and miR-19-43966-3p were detected in mRNA of *SCAP* gene forming a cluster of 39 nt in length from 96 to 135 nt. Without overlapping sites, their length would be 294 nt, which exceeds the 5'UTR size of 259 nt.

**Table 1** – Characteristics of miRNAs interaction in the 5'UTR of mRNA of metabolic syndrome candidate gene

Gene	miRNA	Position, nt	$\Delta G$ , kJ/mole	$\Delta G/\Delta G_m$ , %	Length, nt
<i>ACSL1</i>	miR-19-42639-3p	81	-115	90	22
<i>ACVR1C</i>	miR-5-15435-5p	9	-108	93	20
	miR-22-46172-3p	207	-121	90	22
<i>ADRA2A</i>	miR-2-3578-5p	36	-110	96	18
	miR-17-38379-3p	496	-119	89	23
	miR-11-29838-3p	871	-121	89	23
	miR-4-13479-5p	913	-106	93	20
<i>ALDH2</i>	miR-21-16482-3p	8	-123	92	21
<i>APLN</i>	miR-6-16211-3p	263	-123	89	23
<i>AR</i>	miR-19-41434-3p	621	-115	95	21
<i>BRAP</i>	miR-X-44865-3p	162	-115	92	20
<i>CAPN10</i>	miR-6-16793-3p	105	-115	95	20
<i>CAV1</i>	miR-19-41383-3p	311	-115	90	23
	miR-16-39450-3p	779	-117	89	23
<i>CXCL16</i>	miR-2-7122-3p	88	-119	93	21
	miR-17-35758-5p	439	-117	93	22
<i>EPO</i>	miR-12-31979-3p	12	-121	89	23
<i>ESR2</i>	miR-17-38870-3p	41	-117	92	22
<i>FAAH</i>	miR-5-15548-3p	19	-129	92	23
<i>FGF19</i>	miR-12-31701-3p	151	-115	90	22
	miR-11-28201-3p	163	-127	88	24
	miR-13-32816-5p	298	-115	90	22
<i>H6PD</i>	miR-6-16847-3p	58	-121	92	22
<i>HMGAI</i>	miR-22-46516-3p	350	-117	92	21
<i>HMOX1</i>	miR-16-22443-3p	75	-113	95	20

Continuation of table 1

Gene	miRNA	Position, nt	$\Delta G$ , kJ/mole	$\Delta G/\Delta G_m$ , %	Length, nt
<i>HNF4A</i>	miR-9-26042-5p	53	-125	92	22
<i>HSD11B1</i>	miR-9-26506-3p	24	-115	93	22
<i>HTR2A</i>	miR-1-2136-3p	600	-104	91	21
<i>HTR2C</i>	miR-4-10151-3p	424	-119	90	22
<i>ICAM1</i>	miR-1-1109-3p	106	-117	89	23
<i>IGFBP2</i>	miR-17-39570-5p	65	-123	91	22
	miR-19-34424-3p	84	-113	95	20
	miR-8-22971-3p	86	-117	92	21
<i>IL6R</i>	miR-19-30988-5p	329	-129	90	23
	miR-10-13751-3p	336	-121	92	21
	miR-7-15849-3p	341	-110	96	18
<i>INPPL1</i>	miR-3-7790-3p	96	-121	93	21
<i>INSIG1</i>	miR-4-11181-3p	5	-115	93	20
	miR-11-14307-3p	53	-115	92	21
<i>MMP2</i>	miR-2-6328-5p	264	-117	89	23
<i>MTMR9</i>	miR-18-41332-3p	281	-125	91	23
	miR-2-6530-5p	288	-119	92	22
	miR-2-5888-3p	290	-125	91	23
<i>NEDD4L</i>	miR-5-12460-5p	56	-129	90	24
<i>NOS3</i>	miR-3-8846-5p	200	-123	88	24
<i>NPY2R</i>	miR-7-20689-3p	903	-96	92	20
<i>PDK4</i>	miR-11-12657-3p	47	-108	91	21
	miR-19-34424-3p	216	-110	93	20
<i>PLTP</i>	miR-17-12804-3p	108	-113	93	20
<i>PTEN</i>	miR-20-43459-5p	75	-115	92	20
	miR-5-15564-3p	486	-125	91	22
	miR-17-41310-3p	708	-110	96	18
	miR-3-9461-3p	790	-121	89	23
<i>SCAP</i>	miR-12-30825-5p	22	-115	92	22
	miR-12-31721-3p	23	-108	91	21
<i>SERPINE1</i>	miR-16-38458-3p	30	-123	88	24
<i>SH2B1</i>	miR-11-23098-5p	225	-110	91	21
<i>STEAP4</i>	miR-14-33186-5p	49	-129	88	24
<i>TGFB1</i>	miR-20-43381-5p	1	-121	92	21
	miR-5-8853-5p	6	-115	92	20
	miR-9-13610-3p	6	-121	92	21
	miR-12-30416-5p	186	-117	92	22
	miR-10-13655-3p	209	-129	95	22
	miR-11-29785-3p	232	-108	91	21
	miR-11-29785-5p	232	-108	91	21
	miR-9-26506-3p	237	-113	91	22
<i>TPM1</i>	miR-17-38733-3p	241	-119	89	24
	miR-19-8151-3p	168	-117	92	21

Continuation of table 1

Gene	miRNA	Position, nt	$\Delta G$ , kJ/mole	$\Delta G/\Delta G_m$ , %	Length, nt
<i>TRIB1</i>	miR-2-3313-3p	233	-142	89	25
	miR-17-40348-5p	239	-127	94	23
	miR-1-155-3p	245	-127	92	22
	miR-2-7252-3p	303	-123	88	24
	miR-4-7472-5p	360	-110	91	21

Binding sites for five miRNAs were identified in mRNA of *MMP2* gene in the region from 110 nt to 125 nt, the miRNAs lengths without sites overlapping would be 114 nt.

Four binding sites in the mRNA of *TGFB1* gene formed a 33 nt long cluster from 232 nt to 265 nt, without overlapping miRNAs lengths would be equal to 88 nt.

mRNAs of *MTMR9* and *TRIB1* genes had three miRNA binding sites formed in the cluster. Four miRNA binding sites were found in mRNAs of *PTEN* and *TGFB1* genes in the cluster from 531 to 558 nt and from 232 to 265, respectively. In the mRNA of *PNPLA3* gene, binding sites of miR-X-13195-3p, miR-1-265-3p, miR-19-33623-3p, miR-17-41168-3p, and miR-1-1714-3p are located in a

cluster from 148 nt to 172 nt, the total length of the miRNAs is 109 nt.

The average free energy of miRNAs binding with all target mRNAs in 5'UTR was equal to  $-122,4 \pm -10,5$  kJ/mole. There were 69 associations of miRNAs with mRNAs having a free energy of binding of more than -120 kJ/mole.

The following miRNAs and gene pairs were established as effective associations: miR-2-3313-3p and *TRIB1*, miR-2-3313-3p and *SCAP*, miR-1-2121-3p and *SCAP*, miR-19-21199-3p and *SCAP*, miR-1-2121-3p and *ADRA2A*, miR-19-21199-3p and *ADRA2A* with free interaction energy of -140 kJ/mole and more.

Of the 66 miRNAs target genes in CDS, their mRNA was associated with 133 miRNAs (Table 2).

**Table 2** – Characteristics of miRNAs interaction in the CDS of mRNA of metabolic syndrome candidate gene

Gene	miRNA	Position, nt	$\Delta G$ , kJ/mole	$\Delta G/\Delta G_m$ , %	Length, nt
<i>ABCA1</i>	miR-11-21109-3p	6416	-110	90	23
<i>ACE</i>	miR-10-27065-3p	60	-117	93	21
	miR-11-28656-5p	62	-125	89	23
	miR-2-3313-3p	63	-138	87	25
	miR-3-8100-5p	64	-132	90	24
<i>ACVR1C</i>	miR-17-39672-3p	278	-113	91	21
<i>ADIPOR1</i>	miR-6-16717-3p	1092	-121	93	22
<i>ADRA1A</i>	miR-17-34996-5p	1761	-113	91	23
<i>ADRA2A</i>	miR-7-20411-3p	1064	-123	91	23
	miR-X-44737-3p	1526	-123	89	24
	miR-19-43342-3p	1535	-119	90	22
	miR-5-15733-3p	1536	-132	89	24
	miR-16-38088-5p	1842	-123	89	23
	miR-13-32613-3p	2006	-125	88	24
	miR-19-43736-3p	2049	-119	90	22
<i>ADRA2B</i>	miR-15-36451-5p	455	-121	89	23
	miR-22-44023-3p	465	-121	92	21
	miR-12-32603-3p	882	-115	92	23
	miR-19-43065-3p	889	-113	90	22

Continuation of table 2

Gene	miRNA	Position, nt	$\Delta G$ , kJ/mole	$\Delta G/\Delta G_m$ , %	Length, nt
<i>ADRB3</i>	miR-21-45808-3p	1138	-115	95	20
	miR-2-4736-5p	1306	-121	92	21
	miR-1-654-3p	1312	-115	92	20
	miR-5-8853-5p	1312	-115	92	20
<i>AGTR1</i>	miR-6-3109-5p	102	-117	92	22
<i>ALDH2</i>	miR-1-3575-5p	1559	-119	89	23
<i>ANGPTL4</i>	miR-19-43315-5p(2)	258÷259	-121÷-134	90÷100	23
	miR-9-26025-3p	567	-113	90	22
<i>ANGPTL6</i>	miR-19-43342-3p	433	-121	92	22
	miR-7-20142-5p	526	-119	89	23
	miR-19-42224-5p	535	-117	95	21
	miR-6-18047-5p	599	-100	94	19
	miR-16-37839-3p	855	-117	90	23
<i>APOA1</i>	miR-10-13655-3p	841	-123	91	22
<i>APOB</i>	miR-13-36375-5p	179	-119	90	23
	miR-19-25731-3p	2054	-93	92	20
	miR-19-25731-5p	2054	-93	92	20
<i>APOE</i>	miR-X-45440-5p	643	-121	95	22
	miR-9-23547-5p	766	-115	93	20
	miR-9-24355-5p	768	-115	93	20
<i>AR</i>	miR-10-26714-5p	1363	-127	94	24
	miR-17-35260-3p	1517	-115	90	22
	miR-17-40389-5p	1744	-113	95	20
	miR-5-15733-3p	2514	-134	90	24
	miR-1-1819-3p(2)	2515÷2518	-123÷-125	89÷91	23
<i>BTN2A1</i>	miR-10-26815-5p	280	-121	88	24
<i>CAPN10</i>	miR-22-45441-3p	474	-115	89	23
	miR-2-5634-5p	1114	-104	89	24
<i>CDH13</i>	miR-2-7128-3p	1089	-102	89	23
<i>CEBPA</i>	miR-19-28028-5p	233	-132	89	24
	miR-21-23994-3p	236	-113	91	21
	miR-20-40417-3p	777	-113	96	19
	miR-22-16963-5p	782	-127	91	22
	miR-6-16525-3p	885	-119	89	23
<i>CLOCK</i>	miR-3-11123-5p	1181	-98	90	22
<i>CUL7</i>	miR-11-1939-5p	124	-115	92	20
	miR-19-42999-3p	832	-110	90	22
	miR-22-45904-3p	3313	-115	92	22
	miR-17-38391-3p	4439	-119	93	23
	miR-13-35476-3p	4443	-117	90	22
	miR-9-25099-3p	4444	-115	90	22
<i>CYP46A1</i>	miR-1-527-3p	1469	-106	93	20
<i>DYRK1B</i>	miR-5-15058-5p	1562	-113	93	20
	miR-9-25558-3p	1810	-125	88	24

Continuation of table 2

Gene	miRNA	Position, nt	$\Delta G$ , kJ/mole	$\Delta G/\Delta G_m$ , %	Length, nt
<i>ENPP1</i>	miR-X-44972-5p	32	-119	93	20
<i>EPO</i>	miR-3-8171-3p	741	-110	93	22
<i>ESR1</i>	miR-5-4100-5p	409	-106	91	22
	miR-4-5310-3p	1852	-115	90	23
<i>FAAH</i>	miR-17-38856-3p	108	-115	93	20
	miR-15-36321-3p	901	-115	90	22
<i>FADS2</i>	miR-16-33426-5p	1447	-110	90	22
<i>FGF23</i>	miR-2-7679-5p	301	-102	91	22
<i>GCKR</i>	miR-10-26109-5p	820	-119	92	22
<i>GPX1</i>	miR-5-15733-3p	98	-134	90	24
	miR-5-16634-3p	338	-121	90	22
<i>H6PD</i>	miR-4-12982-5p	519	-110	91	21
	miR-7-20135-3p	1080	-110	90	22
<i>HMGAI</i>	miR-19-43704-3p	550	-115	90	23
<i>HNF1A</i>	miR-4-12346-5p	1463	-108	89	23
	miR-8-24124-3p	1825	-113	90	22
<i>HP</i>	miR-1-3943-5p	1178	-98	92	20
<i>IGFBP2</i>	miR-15-39164-3p	214	-121	97	20
<i>IL6R</i>	miR-2-4533-3p	483	-125	89	23
<i>INPPL1</i>	miR-5-14202-5p	161	-123	91	22
	miR-20-42676-3p	161	-127	91	23
	miR-22-45334-5p	169	-123	92	23
	miR-10-26505-5p	1162	-119	89	23
<i>INS</i>	miR-16-38416-3p	402	-115	90	22
<i>INSIG1</i>	miR-8-23775-5p	339	-117	95	21
	miR-14-14807-5p	414	-110	91	21
	miR-11-28905-3p	601	-117	89	23
<i>INSR</i>	miR-4-11316-5p	127	-132	89	24
<i>IRS1</i>	miR-3-7886-3p	412	-127	88	24
	miR-8-23997-5p	1418	-102	94	19
	miR-22-45452-5p	2084	-102	92	20
	miR-10-26714-5p	2085	-121	89	24
	miR-19-37450-3p	3446	-106	93	21
<i>KL</i>	miR-1-3822-3p	539	-121	92	22
	miR-19-41858-5p	2542	-121	89	23
<i>LDLR</i>	miR-10-26537-5p	2452	-108	96	20
<i>LMNA</i>	miR-16-37015-3p	1392	-113	93	20
<i>LRP5</i>	miR-17-38580-3p	148	-115	90	22
	miR-2-2243-3p	2090	-106	91	21
	miR-19-41684-3p	2469	-121	89	24
	miR-1-3919-5p	4788	-121	88	24
	miR-19-38895-3p	4824	-123	89	24
<i>LRP6</i>	miR-7-20135-3p	3421	-110	90	22

Continuation of table 2

Gene	miRNA	Position, nt	$\Delta G$ , kJ/mole	$\Delta G/\Delta G_m$ , %	Length, nt
<i>MMP2</i>	miR-21-45324-5p	379	-125	91	23
	miR-19-43421-5p	1681	-108	91	21
	miR-17-39037-3p	1691	-113	90	22
<i>MMP9</i>	miR-17-38947-5p(2)	219÷220	-113	90	22
	miR-16-34158-5p	698	-113	93	22
<i>MTRR</i>	miR-12-31626-5p	1751	-108	89	23
<i>NEDD4L</i>	miR-5-14114-5p	853	-123	89	23
	miR-3-9322-5p	2585	-102	91	21
<i>NOS3</i>	miR-15-38767-3p	2946	-123	89	24
	miR-X-45814-5p	3073	-117	89	24
	miR-19-43338-3p	3599	-117	90	22
<i>PNPLA3</i>	miR-22-46211-3p	918	-106	93	22
<i>PRDM16</i>	miR-4-11714-5p	2235	-110	96	20
	miR-11-27530-5p	2876	-119	89	23
	miR-11-29785-3p	3424	-108	91	21
	miR-11-29785-5p	3424	-108	91	21
<i>RLN3</i>	miR-16-33426-5p	46	-110	90	22
	miR-17-39466-3p	55	-110	90	22
	miR-3-7740-5p	304	-113	96	20
<i>SCAP</i>	miR-12-17092-3p	2486	-125	91	22
<i>SERPINE1</i>	miR-2-3962-5p	542	-125	88	24
<i>SH2B1</i>	miR-5-15578-5p	2148	-119	89	23
	miR-4-13219-5p	2834	-106	91	22
	miR-1-1109-3p	2898	-117	89	23
<i>SIRT1</i>	miR-5-13181-3p	232	-123	89	24
	miR-9-25099-3p	435	-115	90	22
<i>SREBF2</i>	miR-1-2002-3p	564	-121	90	22
<i>TNF</i>	miR-20-42898-3p	230	-121	92	23
	miR-20-42898-5p	230	-121	92	23
<i>TPM1</i>	miR-15-35627-5p(2)	332÷333	-117÷-123	95÷100	22
<i>TRIB1</i>	miR-8-24549-5p	756	-127	90	24
<i>UCP3</i>	miR-19-42357-3p	406	-115	89	23
<i>VEGFA</i>	miR-9-26506-3p	775	-113	91	22
	miR-8-21883-3p	887	-123	88	24

mRNA of *AR* gene contained 21 miRNA binding sites for miR-9-20317-3p and miR-17-39416-3p, whose nucleotide sequences are overlapped.

miR-10-27065-3p, miR-11-28656-5p, miR-2-3313-3p, miR-3-8100-5p binding sites were identified in a cluster in the region from 60 to 88 nt of mRNA of *ACE* gene, the total length of the miRNAs is 93 nt.

The 22 miRNA binding sites in mRNA of *CEBPA* gene formed a cluster with a length of 35

nt, which, without overlapping of sites, would be 308 nt.

18 binding sites for 11 miRNAs were identified in mRNA of *IGFBP2* gene in the region from 139 nt to 177 nt.

There is a 45 nt in length cluster in mRNA of *KL* gene formed by 14 binding sites of 11 miRNAs with a total length of 279 nt, therefore such a compact site arrangement is necessary to reduce the proportion of binding sites.

Six binding sites in mRNA of *SIRT1* gene formed a 60 nt long cluster from 264 nt to 324 nt, without overlapping miRNAs lengths is equal to 142 nt. mRNAs of *CUL7* and *INPPL1* genes contained 3 binding sites of miRNAs, forming a cluster.

The average free energy of binding of miRNAs with all mRNAs in CDS is equal to  $-118 \pm 9$  kJ/mole. There are 120 associations of miRNAs with mRNAs that have a free energy of binding of more than -120 kJ/mole.

50 target genes were associated with 82 miRNAs in 3'UTR (Table 3).

Five miRNAs had binding sites in mRNA of *CD36* gene, forming a cluster with a length of 40 nt.

The total length of these miRNAs was 115 nt, which is almost 50% of the 275-nt CDS length. mRNA of *IGF1* gene contains nine miRNA-binding sites for miR-3-5147-5p and miR-101-27078-5p, which nucleotide sequences completely coincided, with a cluster length of 39 nt. mRNA of *JAK2* gene bound with miR-10-29282-3p (ten binding sites) and miR-15-36862-3p (nine binding sites) and the formed clusters coincide.

mRNAs of *CD36*, *IGF1* and *JAK2* genes contained 15, 18 and 19 binding sites, respectively, which miRNA nucleotide sequences overlapped. Four miRNA binding sites formed a cluster with a length of 67 nt from 7321 nt to 7388 nt in mRNA of *IRS1* gene.

**Table 3 – Characteristics of miRNAs interaction in 3'UTR of mRNA of metabolic syndrome candidate gene**

Gene	miRNA	Position, nt	$\Delta G$ , kJ/mole	$\Delta G/\Delta G_m$ , %	Length, nt
<i>ACE</i>	miR-13-28252-3p	4068	-117	90	22
	miR-1-2030-3p	4657	-110	90	22
	miR-X-46577-3p	4792	-106	91	21
<i>ADIPOQ</i>	miR-17-39935-3p	1651	-104	91	21
<i>ADRA1A</i>	miR-6-19858-3p	1975	-108	91	22
<i>ADRB3</i>	miR-10-26254-3p	2347	-121	88	24
	miR-6-17487-3p	2442	-115	92	23
	miR-10-29282-3p	2456	-108	93	23
	miR-15-36862-3p	2456	-113	93	23
	miR-15-36862-3p	2462	-113	93	23
<i>AGTR2</i>	miR-16-40163-5p	2307	-121	90	23
<i>AHII</i>	miR-22-45902-3p	4645	-113	93	22
	miR-9-21385-3p	5491	-96	90	22
<i>AKT1</i>	miR-10-27065-3p	2864	-117	93	21
	miR-12-5800-5p	2866	-113	93	20
	miR-13-36375-5p	2875	-119	90	23
<i>ANGPT2</i>	miR-7-21133-5p	3064	-121	89	24
	miR-5-18072-3p	3071	-102	91	22
<i>AR</i>	miR-3-10752-3p	4096	-108	89	23
<i>CD36</i>	miR-12-31413-3p	3526	-104	89	23
	miR-15-36862-3p(5)	3529÷3539	-108	89	23
	miR-10-29282-3p(5)	3533÷3539	-104	89	23
	miR-19-42814-5p	3542	-106	91	23
	miR-10-29282-3p	3543	-104	89	23
<i>CEBPA</i>	miR-22-45967-3p	2440	-115	92	22
<i>CYP11B2</i>	miR-17-12514-5p	1982	-104	92	20
<i>CYP46A1</i>	miR-13-33774-5p	1571	-123	88	24

Continuation of table 3

Gene	miRNA	Position, nt	$\Delta G$ , kJ/mole	$\Delta G/\Delta G_m$ , %	Length, nt
<i>DIO2</i>	miR-11-28228-3p	4777	-96	92	20
	miR-11-27435-3p	6187	-96	90	22
<i>DYRK1B</i>	miR-11-28385-3p	2463	-104	92	20
<i>ENPP1</i>	miR-10-26483-5p	6274	-110	90	22
	miR-2-4804-5p(2)	6751÷6752	-113÷117	90÷93	24
<i>ESR1</i>	miR-8-24024-3p	3339	-121	88	24
<i>FADS2</i>	miR-19-43386-3p	2407	-117	89	23
	miR-1-1412-5p	2712	-117	90	22
	miR-17-39583-3p	2803	-119	89	23
<i>FTO</i>	miR-17-34996-5p	3710	-110	90	23
	miR-2-5355-3p	3906	-115	90	22
<i>GCKR</i>	miR-6-18764-3p	1956	-121	88	24
<i>H6PD</i>	miR-1-318-5p	3669	-115	92	22
	miR-22-45335-5p	5829	-113	90	23
	miR-X-46030-5p	6616	-110	90	22
<i>HMGAI</i>	miR-15-38620-5p	877	-119	90	22
<i>ICAMI</i>	miR-15-36862-3p	2987	-108	89	23
	miR-17-39935-3p	3022	-104	91	21
	miR-10-26483-5p	3025	-110	90	22
<i>IGF1</i>	miR-3-5147-5p(9)	4042÷4058	-100	90	22
	miR-101-27078-5p(9)	4042÷4058	-108	89	23
<i>IGF2BP3</i>	miR-X-48172-3p	4088	-104	91	22
<i>IL10</i>	miR-17-39466-3p	1200	-110	90	22
<i>IL6R</i>	miR-14-35161-5p	3063	-119	90	24
<i>INPPL1</i>	miR-17-40267-5p	4336	-125	88	24
	miR-10-28609-3p	4512	-104	91	22
<i>INSR</i>	miR-19-42303-3p	5363	-117	90	23
	miR-12-30825-5p	5366	-115	92	22
	miR-12-31721-3p	5367	-115	96	21
<i>IRSI</i>	miR-10-29282-3p(2)	7402÷7404	-104	89	23
	miR-10-29282-3p	7425	-104	89	23
<i>JAK2</i>	miR-10-29282-3p(10)	5184÷5202	-104÷108	89÷91	23
	miR-15-36862-3p(9)	5184÷5200	-108	89	23
<i>LCN2</i>	miR-22-23987-3p	682	-123	94	21
<i>LDLR</i>	miR-17-39466-3p	3887	-110	90	22
	miR-8-11096-5p	3890	-113	90	22
	miR-X-45975-5p	4004	-96	92	22
	miR-4-12245-3p(2)	4559÷4560	-110	90	22
	miR-2-4826-5p(2)	4607÷4608	-113÷115	90÷92	23
	miR-7-20771-3p	4974	-89	91	21
<i>LEP</i>	miR-8-11096-5p	3087	-113	90	22
	miR-17-35758-5p	3092	-113	90	22

Continuation of table 3

Gene	miRNA	Position, nt	$\Delta G$ , kJ/mole	$\Delta G/\Delta G_m$ , %	Length, nt
<i>LMNA</i>	miR-18-41332-3p	2808	-123	89	23
	miR-11-28484-5p	2820	-117	90	22
	miR-19-42375-3p	3033	-113	93	21
<i>LPIN1</i>	miR-17-40078-3p	4485	-113	88	24
<i>LTA</i>	miR-16-9117-3p	1258	-98	92	21
<i>MTHFR</i>	miR-9-24450-5p	3350	-117	89	24
	miR-10-11641-3p	6281	-119	89	23
	miR-X-44909-3p	6342	-108	91	22
	miR-2-4684-5p	6844	-117	93	22
	miR-22-45902-3p	7051	-113	93	22
<i>MTMR9</i>	miR-17-8001-3p	4094	-110	90	23
	miR-X-45975-5p	6064	-96	92	22
	miR-2-4826-5p(2)	6186÷6187	-113	90	23
<i>MTTP</i>	miR-19-42953-5p	3438	-113	91	22
<i>NPY2R</i>	miR-3-9607-3p	2968	-96	90	22
<i>OLR1</i>	miR-3-5147-5p(2)	1504÷1506	-100	90	22
	miR-101-27078-5p(2)	1504÷1506	-108	89	23
<i>PRDM16</i>	miR-19-43175-3p	4675	-113	91	21
	miR-22-44124-5p	4961	-115	89	23
	miR-8-23404-5p	5193	-125	88	24
	miR-X-38664-5p	5199	-110	93	22
<i>PYY</i>	miR-1-252-5p	489	-106	91	21
<i>SH2B1</i>	miR-16-40163-5p	4530	-119	89	23
<i>SLC22A12</i>	miR-7-17280-5p	2874	-119	90	22
<i>SREBF2</i>	miR-17-38738-5p	4196	-117	90	22
	miR-1-2142-3p	4206	-123	92	23
<i>STEAP4</i>	miR-17-34996-5p	3201	-113	91	23
<i>TGFB1</i>	miR-9-13610-3p	2060	-123	94	21
	miR-17-12804-3p	2062	-113	93	20
	miR-8-24549-5p	2066	-125	88	24
	miR-15-38620-5p	2089	-119	90	22
	mir-1-2121-3p	2093	-140	89	25
<i>TRIB3</i>	miR-4-5601-5p	1228	-117	90	22
	miR-16-37914-3p	1540	-121	88	25

Five miRNAs binding sites were found in mRNA of *TGFB1* gene, from 2060 nt to 2118 nt, forming a cluster with a length of 58 nt, considering a total length of miRNAs sequences is 112. nt

mRNAs of *AKT1* and *TGFB1* genes had three miRNA binding sites formed into clusters, and the mRNA of *ADRB3* gene contains the binding sites of 4 miRNAs from 2442 to 2485 forming the 43 nt

length cluster while a total length of the miRNAs sequences is 92 nt.

The average free energy of binding of miRNAs with all mRNAs in 3'UTR was -111 nt. 15 associations of miRNAs and mRNAs were found with a free binding energy of more than -120 kJ/mole.

Among 109 genes participating in the development of MS, 41 genes were associated with

70 miRNAs in 5'UTR, 66 genes with 133 miRNAs in CDS, and 50 genes with 82 miRNAs in 3'UTR.

Consequently, almost half of the miRNAs are bound at the beginning of the nucleotide sequence of mRNA (in the 5'UTR), which makes it possible to stop protein synthesis before the translation stage. This allows you to save energy resources of the cell, since stopping protein synthesis at the late stage of translation may result in the termination of protein synthesis with the formation of defective polypeptides. Binding of miRNA to mRNA at the beginning of the CDS will also help save energy resources. Therefore, the majority of miRNA binding sites in the mRNA of candidate MS genes are located in the CDS immediately after 5'UTR. In addition to the preferential localization of miRNA binding sites at the beginning of mRNA, the free energy of miRNA interaction with mRNA plays an important role in the process of regulating gene expression. As a rule, the free energy of the interaction of miRNA with mRNA for binding sites located in 5'UTR is greater than for binding sites located in CDS. And the free energy of miRNA interaction with mRNA for binding sites in CDS is greater than for binding sites located in 3'UTR.

Of them, 27 genes that bound to only one miRNA were located in 5'UTR region, 33 genes in CDS, and 23 genes in 3'UTR. The remaining mRNAs of genes involved in the development of MS were associated with two or more miRNAs. Thus, 14 genes had miRNA binding sites in 5'UTR, 33 genes in CDS, and 27 in 3'UTR. Consequently, most of miRNA-mRNA associations are located in CDS. mRNAs of following genes interacted with miRNAs with the highest value of free binding energy: *SCAP* gene (ten miRNAs) in 5'UTR, *CEBPA* gene in CDS (seven miRNAs), and *TGFB1* gene in 3'UTR (one miRNAs). The binding sites located in 5'UTR and

CDS had the strongest interaction. Only *H6PD*, *HMGAI*, *IL6R* and *INPPL1* genes had binding sites in 5'UTR, CDS and 3'UTR. The presence of many binding sites for several miRNAs in mRNA genes indicates a strong dependence of their expression on miRNA.

Identified in the mRNA of some genes, the binding sites of two or more miRNAs, with overlapping nucleotide sequences, lead to competition between these miRNAs for binding to mRNA. The overlap of nucleotide sequences of binding sites reduces their share in the total length of mRNA.

## Conclusion

It was established that the genes responsible for the development of metabolic syndrome are regulated by miRNAs. So *ADRA2A*, *PTEN*, *SCAP* and *TGFB1* genes are regulated by binding of 12, 9, 15 and 8 miRNAs to 5'UTR with the highest binding energy is equal to -144 kJ/mole, -132 kJ/mole, -151 kJ/mole and -129 kJ/mole, respectively. *APRA2A*, *AR*, *CEBPA*, *IGFBP2*, *KL* and *SIRT1* genes are regulated by 7, 7, 19, 11, 13 and 6 miRNAs binding to CDS with the highest binding energy being -132 kJ/mole, -134 kJ/mole, -142 kJ/mole, -140 kJ/mole, -142 kJ/mole and -138 kJ/mole, respectively. *ADRA2A*, *SCAP*, *AR*, *CEBPA*, *IGFBP2*, *KL*, and *SIRT1* genes had multiple binding sites of miRNAs, forming clusters. Thus, these genes can be used as potential markers for the diagnosis of the metabolic syndrome.

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## References

- 1 Lupton SJ., Chiu CL., Lind JM. A hypertension gene: are we there yet? // Twin Res Hum Genet. -2011. – Vol. 14, No.4. – P.295-304. doi: 10.1375/twin.14.4.295.
- 2 Stančáková A., Laakso M. Genetics of metabolic syndrome // Rev Endocr Metab Disord. – 2014. – Vol. 15, No.4. – P.243-252. doi: 10.1007/s11154-014-9293-9.
- 3 Aleksandrova K., Boeing H., Jenab M., et al. Metabolic syndrome and risks of colon and rectal cancer: the European prospective investigation into cancer and nutrition study // Cancer Prev. Res. – 2011. – Vol. 4. – P. 1873–1883.
- 4 Ambler SK., Brown RD. Genetic determinants of blood pressure regulation // J Cardiovasc Nurs. – 1999. – Vol. 13, No.4. – P.59-77.
- 5 Bochud M., Bovet P., Burnier M., Eap CB. CYP3A5 and ABCB1 genes and hypertension // Pharmacogenomics. – 2009. – Vol. 10, No.3. – P.477-487. doi: 10.2217/14622416.10.3.477.

- 6 Gatti RR., Santos PS., Sena AA., Marangoni K., Araújo MA., Goulart LR. The interaction of AGT and NOS3 gene polymorphisms with conventional risk factors increases predisposition to hypertension // *J Renin Angiotensin Aldosterone Syst.* – 2013. – Vol. 14, No.4. – P.360-368. doi: 10.1177/1470320312452027.
- 7 Chen K., Fu C., Chen C., et al Role of GRK4 in the regulation of arterial AT1 receptor in hypertension // *Hypertension*. – 2014. – Vol. 63, No.2. – P.289-296. doi: 10.1161/HYPERTENSIONAHA.113.01766.
- 8 Vargas T., Moreno-Rubio J., Herranz J. Genes associated with metabolic syndrome predict disease-free survival in stage II colorectal cancer patients. A novel link between metabolic dysregulation and colorectal cancer // *Mol Oncol.* – 2014. – Vol. 8, No.8. – P.1469-1481. doi: 10.1016/j.molonc.2014.05.015.
- 9 Hagiwara S., Kantharidis P., Cooper ME. MicroRNA as biomarkers and regulator of cardiovascular development and disease // *Curr Pharm Des.* – 2014. – Vol. 20, No.14. – P.2347-2370.
- 10 Yuan LQ., de Jesus Perez V., Liao XB., Król M., Yeh CH. MicroRNA and Cardiovascular Disease 2016 // *Biomed Res Int.* – 2017. – Vol. 2017. – P.3780513. doi: 10.1155/2015/734380.
- 11 Bátka S., Thum T. MicroRNAs in hypertension: mechanisms and therapeutic targets // *Curr Hypertens Rep.* – 2012. – Vol. 14, No.1. – P.79-87. doi: 10.1007/s11906-011-0235-6.
- 12 Zhongguo Zhong Yao Za Zhi. MicroRNA and hypertension // Article in Chinese. – 2014. – Vol. 39, No.3. – P.397-401;
- 13 Meloche J., Paulin R., Provencher S., Bonnet S. Therapeutic Potential of microRNA Modulation in Pulmonary Arterial Hypertension // *Curr Vasc Pharmacol.* – 2015. – Vol. 13, No.3. – P.331-340.
- 14 Zhou G., Chen T., Raj JU. MicroRNAs in pulmonary arterial hypertension // *Am J Respir Cell Mol Biol.* – 2015. – Vol. – 52, No.2. – P.139-151. doi: 10.1165/rcmb.2014-0166TR.
- 15 Synetos A., Toutouzas K., Stathogiannis K., Latsios G., Tsiamis E., Tousoulis D., Stefanadis C. MicroRNAs in arterial hypertension // *Curr Top Med Chem.* – 2013. – Vol. 13, No.13. – P.1527-1532.
- 16 Boucherat O., Potus F., Bonnet S. microRNA and Pulmonary Hypertension // *Adv Exp Med Biol.* – 2015. – Vol. 888. – P.237-252. doi: 10.1007/978-3-319-22671-2\_12.
- 17 Bienertova-Vasku J., Novak J., Vasku A. MicroRNAs in pulmonary arterial hypertension: pathogenesis, diagnosis and treatment // *J Am Soc Hypertens.* – 2015. – Vol. 9, No.3. – P.221-234. doi: 10.1016/j.jash.2014.12.011.
- 18 Lee A., McLean D., Choi J., Kang H., Chang W., Kim J. Therapeutic implications of microRNAs in pulmonary arterial hypertension // *BMB Rep.* – 2014. – Vol. 47, No.6. – P.311-317.
- 19 Karolina DS., Tavintharan S., Armugam A., et al Circulating miRNA profiles in patients with metabolic syndrome // *J Clin Endocrinol Metab.* – 2012. – Vol. 97, No.12 – P.2271-2276. doi: 10.1210/jc.2012-1996.
- 20 Price NL., Ramírez CM., Fernández-Hernando C. Relevance of microRNA in metabolic diseases // *Crit Rev Clin Lab Sci.* – 2014. – Vol. 51, No.6. – P.305-320. doi: 10.3109/10408363.2014.937522.
- 21 Ramírez CM., Goedeke L., Fernández-Hernando C. “Micromanaging” metabolic syndrome // *Cell Cycle.* – 2011. – Vol. 10, No.19. – P.3249-3252. doi: 10.4161/cc.10.19.17558.
- 22 Londin E., Lohera P., Telonisa A.G., Quanna K. et al. Analysis of 13 cell types reveals evidence for the expression of numerous novel primate and tissue-specific microRNAs // *PNAS USA.* – 2015. – Vol.112. – P.1106-1115. doi: 10.1073/pnas.1420955112.
- 23 Ivashchenko A.T., Pyrkova A.Y., Niyazova R.Y., Alybayeva A., Baskakov K. Prediction of miRNA binding sites in mRNA // *Bioinformation.* – 2016. – Vol.12. – P.237-240.
- 24 Kool E.T. Hydrogen bonding, base stacking, and steric effects in DNA replication // *Annual Review of Biophysics and Biomolecular Structure.* – 2001. – Vol.30. P.1–22. doi:10.1146/annurev.biophys.30.1.1.
- 25 Leontis N.B., Stombaugh J., Westhof E. The non-Watson- Crick base pairs and their associated isostericity matrices // *Nucleic Acids Research.* – 2002. – Vol.30. P.3497–3531.

## References

- 1 Aleksandrova K., Boeing H., Jenab M., et al. (2011) Metabolic syndrome and risks of colon and rectal cancer: the European prospective investigation into cancer and nutrition study. *Cancer Prev. Res.*, vol. 4, p. 1873–1883.
- 2 Ambler SK., Brown RD. (1999) Genetic determinants of blood pressure regulation. *J Cardiovasc Nurs.*, vol. 13(4), p.59-77.
- 3 Bátka S., Thum T. (2012) MicroRNAs in hypertension: mechanisms and therapeutic targets. *Curr Hypertens Rep.*, vol. 14(1), p.79-87. doi: 10.1007/s11906-011-0235-6.
- 4 Bienertova-Vasku J., Novak J., Vasku A. (2015) MicroRNAs in pulmonary arterial hypertension: pathogenesis, diagnosis and treatment. *J Am Soc Hypertens.*, vol. 9(3), p.221-234. doi: 10.1016/j.jash.2014.12.011.
- 5 Bochud M., Bovet P., Burnier M., Eap CB. CYP3A5 and ABCB1 genes and hypertension (2009) *Pharmacogenomics*, vol. 10(3), p.477-487. doi: 10.2217/14622416.10.3.477.
- 6 Boucherat O., Potus F., Bonnet S. (2015) microRNA and Pulmonary Hypertension. *Adv Exp Med Biol.*, vol. 888, p.237-252. doi: 10.1007/978-3-319-22671-2\_12.
- 7 Chen K., Fu C., Chen C., et al. (2014) Role of GRK4 in the regulation of arterial AT1 receptor in hypertension. *Hypertension*, vol. 63(2), p.289-296. doi: 10.1161/HYPERTENSIONAHA.113.01766.
- 8 Gatti RR., Santos PS., Sena AA., Marangoni K., Araújo MA., Goulart LR. (2013) The interaction of AGT and NOS3 gene polymorphisms with conventional risk factors increases predisposition to hypertension. *J Renin Angiotensin Aldosterone Syst.*, vol. 14(4), p.360-368. doi: 10.1177/1470320312452027.
- 9 Hagiwara S., Kantharidis P., Cooper ME. (2014) MicroRNA as biomarkers and regulator of cardiovascular development and disease. *Curr Pharm Des.*, vol. 20(14), p.2347-2370.

- 10 Ivashchenko A.T., Pyrkova A.Y., Niyazova R.Y., Alybayeva A., Baskakov K. (2016) Prediction of miRNA binding sites in mRNA. Bioinformation., vol.12, p.237-240.
- 11 Karolina DS., Tavintharan S., Armugam A., et al (2012) Circulating miRNA profiles in patients with metabolic syndrome. *J Clin Endocrinol Metab.*, vol. 97(12), p.2271-2276. doi: 10.1210/jc.2012-1996.
- 12 Kool E.T. (2001) Hydrogen bonding, base stacking, and steric effects in DNA replication. Annual Review of Biophysics and Biomolecular Structure., vol.30, p.1-22. doi:10.1146/annurev.biophys.30.1.1.
- 13 Lee A., McLean D., Choi J., Kang H., Chang W., Kim J. (2014) Therapeutic implications of microRNAs in pulmonary arterial hypertension. *BMB Rep.*, vol. 47(6), p.311-317.
- 14 Leontis N.B., Stombaugh J., Westhof E. (2002) The non-Watson-Crick base pairs and their associated isostericity matrices. *Nucleic Acids Research.*, vol.30, p.3497-3531.
- 15 Londin E., Lohera P., Telonisa A.G., Quanna K. et al. (2015) Analysis of 13 cell types reveals evidence for the expression of numerous novel primate and tissue-specific microRNAs. *PNAS USA.*, vol.112, p.1106-1115. doi: 10.1073/pnas.1420955112.
- 16 Lupton SJ., Chiu CL., Lind JM. (2011) A hypertension gene: are we there yet? *Twin Res Hum Genet.*, vol. 14(4), p.295-304. doi: 10.1375/twin.14.4.295.
- 17 Meloche J., Paulin R., Provencher S., Bonnet S. (2015) Therapeutic Potential of microRNA Modulation in Pulmonary Arterial Hypertension. *Curr Vasc Pharmacol.*, vol. 13(3), p.331-340.
- 18 Price NL., Ramírez CM., Fernández-Hernando C. (2014) Relevance of microRNA in metabolic diseases. *Crit Rev Clin Lab Sci.*, vol. 51(6), p.305-320. doi: 10.3109/10408363.2014.937522.
- 19 Ramírez CM., Goedeke L., Fernández-Hernando C. (2011) "Micromanaging" metabolic syndrome. *Cell Cycle.*, vol. 10(19), p.3249-3252. doi: 10.4161/cc.10.19.17558.
- 20 Stančáková A., Laakso M. (2014) Genetics of metabolic syndrome. *Rev Endocr Metab Disord.*, vol. 15(4), p.243-252. doi: 10.1007/s11154-014-9293-9.
- 21 Synetos A., Toutouzas K., Stathogiannis K., Latsios G., Tsiamis E., Tousoulis D., Stefanadis C. (2013) MicroRNAs in arterial hypertension. *Curr Top Med Chem.* vol. 13(13), p.1527-1532.
- 22 Vargas T., Moreno-Rubio J., Herranz J. (2014) Genes associated with metabolic syndrome predict disease-free survival in stage II colorectal cancer patients. A novel link between metabolic dysregulation and colorectal cancer. *Mol Oncol.*, vol. 8(8), p.1469-1481. doi: 10.1016/j.molonc.2014.05.015.
- 23 Yuan LQ., de Jesus Perez V., Liao XB., Król M., Yeh CH. (2017) MicroRNA and Cardiovascular Disease 2016. *Biomed Res Int.*, vol. 2017, p.3780513. doi: 10.1155/2015/734380.
- 24 Zhongguo Zhong Yao Za Zhi. (2014) MicroRNA and hypertension. Article in Chinese, vol. 39(3), p.397-401;
- 25 Zhou G., Chen T., Raj JU. (2015) MicroRNAs in pulmonary arterial hypertension. *Am J Respir Cell Mol Biol.* vol. – 52(2), p.139-151. doi: 10.1165/rcmb.2014-0166TR.